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Genotype-phenotype correlation in six patients with interstitial deletions spanning 13q31

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4	Genotype-phenotype correlation in six patients with interstitial deletions
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Objective: Since the gene responsible for neurodevelopmental delay observed in patients with chromosomal microdeletions involving the 13q31 region has never been identified, we aimed to narrow the candidate genes. Methods: Analysis of genomic copy number variations was performed as a part of the diagnostic workflow for six patients with neurodevelopmental disorders. Detailed clinical information was collected for each patient carrying a 13q31 deletion. Then, genotype-phenotype correlation analysis was carried out. Results: Genotype-phenotypic correlation analysis excluded regions that were not considered disease-related and focused on the shortest region overlapped between three patients (chr13:75691448_83625667). Among the genes included in this region, MYCBP2 showed a pLI score of "1", suggesting haploinsufficiency intolerance, and was therefore considered as one of the possible candidate genes for neurodevelopmental delay.

Conclusions: There is little information on chromosomal microdeletions involving the
 13q31 region, and more information should be accumulated to better understand the
 genomic contribution of this region.

Keywords: chromosomal microarray testing, genomic copy number variation,

Hirschsprung disease, loss-of-function intolerant

Introduction

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56 Some parts of chromosome 13 display characteristic features. In 1993, Brown et al. reported a clinical case series of patients with 13q microdeletions, and proposed the 57 classification of 13g microdeletions into three groups ^{1,2)}. Group 1: patients with 58 59 proximal deletions not extending into q32, associated with mild or moderate delays in 60 development, with no major anomalies. Group 2: patients with distal deletions, 61 including q32, associated with severe delays in development and major malformations. 62 Group 3: patients with subtelomeric deletions in 13q, associated with severe delays in 63 development, but no major anomalies. The Zic family member 2 gene (ZIC2) located on 64 13q32.3 was later confirmed to be the gene responsible for the main clinical features of group 2, in association with holoprosencephaly ^{3, 4)}. Apart from these classifications, 65 66 deletions in the 13q14 region are associated with retinoblastoma, because the retinoblastoma 1 gene (RB1) responsible for this phenotype is located in this region, and 67 *RB1* haploinsufficiency is causative of retinoblastoma ^{5,6)}. 68 69 Among these subgroups, patients with deletions classified as Groups 1 are rare, and the clinical features are not yet well established ⁷⁻¹⁰. In this study, we aimed to clarify 70 the clinical features of patients with microdeletions involving the 13q31 region, and 71 72 narrow the possible candidate genes for neurodevelopmental delay observed in patients with deletions involving the 13q31 region. 73

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I Materials and methods

Analysis of genomic copy number variations (CNVs) was performed as a part of the diagnostic workflow for patients with neurodevelopmental disorders. This study was performed in accordance with the Declaration of Helsinki, and was approved by the

79 ethics committee of this institution. After receiving written informed consent from 80 patients and their families, we obtained blood samples from the patients, and from their parents when necessary. In this study, genomic copy number analyses were first 81 performed for patients with neurodevelopmental abnormalities of unknown etiology, in 82 accordance with a previously described study 11). Patients with chromosomal 83 84 microdeletions involving the 13q31 region were included in this analysis. Detailed 85 clinical information was collected for each patient carrying a 13q31 deletion. Then, genotype-phenotype correlation analysis was carried out. 86

Genomic positions of CNVs are reported according to the GRCh37/hg19 genome build.

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II Results

1. Chromosomal deletions

We identified six deletions that included the 13q31 region (Figure 1). Genomic positions of the deletions are summarized in Table 1. Because the parents of patient 3 showed no deletion, the deletion identified in patient 3 was confirmed to be de novo.

The deletion identified in patient 6 was inherited from his healthy father, indicating that this deletion was not pathogenic, as reported previously ¹¹⁾. The parents of the other patients declined to be genotyped.

2. Clinical features of patients with microdeletions involving 13q31 region

Clinical information on the patients with microdeletions in the 13q31 region is summarized in Table 1. The details are described below.

A 3-month-old girl (patient 1) was born at term. At birth, the patient was unable to cry. Because vocal cord paralysis was detected by fiberscope, respiratory management

was performed after intubation. Although no clinical findings were suggestive of Hirschsprung disease, weak pigmentation was noted in her iris. She required tube feeding and showed early developmental delay. After tracheostomy, the patient was discharged from the hospital.

A 4 -year-old boy (patient 2) was born at term with normal delivery. At 4 years of age, the patient was examined due to deficient growth, with a height of 88.3 cm (-3.6 SD), a weight of 11.1 kg (-2.6 SD), and an occipitofrontal circumference (OFC) of 47.2 cm (-1.9 SD). However, his growth hormone levels were within normal limits. His development was moderately delayed. He could speak words, but no sentences. He had low-set ears, and micrognathia. No findings were suggestive of Hirschsprung disease or Waardenburg syndrome.

An 11-month-old boy (patient 3) was born at term by normal delivery. Mild right ear deafness (40 dB) and moderate left ear deafness (70 dB) were detected soon after birth, although there was no family history of deafness. The patient showed abnormal pigmentation in his iris, with gray, radial, and sparse pigmentation. A slightly large auricle, wide nasal ala, and decreased muscle tonus (hyperactivity, positive heel-ear sign) were noted. His psychomotor development level was diagnosed as borderline, because he could not move with crawling. At the time of his last hospital visit, his height was 66.0 cm (-1.9 SD), his weight was 7.2 kg (-1.4 SD), and his OFC was 42.0 cm (-2.1 SD), indicating microcephaly.

An 11-year-11-month-old girl (patient 4) was born at 39 weeks of gestation by normal delivery. Her birth weight was 2,284 g (-1.8 SD). Congenital hip dysplasia was noted during infancy. She showed a mild delay in psychomotor development. At the age of 11 years, her height was 136.0 cm (-2.1 SD), her weight was 40.7 kg (standard), and

127 her OFC was 51.7 cm (-0.8 SD), indicating growth deficiency. Her intelligence quotient 128 was evaluated as 60 using the Wechsler Intelligence Scale for ChildrenTM-IV. She 129 underwent precocious puberty, and experienced overactive bladder. Distinctive features 130 were not observed. An 18-month-old girl (patient 5) was born at 39 weeks and 2 days of gestation, with 131 132 a birth weight of 2,114 g (-2.4 SD), a birth length of 44.5 cm (-2.2 SD), and an OFC of 133 33.5 cm (mean). Her mother had no history of miscarriage (she was the first child). 134 Lateral ventricular enlargement was observed by fetal ultrasonography. Brain MRI 135 revealed middle interhemispheric variant of holoprosencephaly ¹²⁾. Distinctive features included narrow palpebral fissures, low-set ears, and a flattened back of the nose. At her 136 137 last visit, she could roll over but could not sit. She could not speak meaningful words. 138 Her length was 74.3 cm (-1.9 SD), her weight was 8.4 kg (-1.5 SD), and her OFC was 40 cm (-4.3 SD), indicating microcephaly. 139 140 A 9-year-old boy (patient 6) was born at 38 weeks of gestation by cesarean section 141 due to fetal head position and pelvic imbalance. Birth weight was 2,556 g (-0.8 SD), 142 birth length was 49 cm (+0.5 SD), and OFC at birth was 33 cm (mean). At present, his 143 height is 112.1 cm (-4.0 SD), his weight is 19.4 kg (-1.8 SD), and his OFC is 51.0 cm (-144 1.3 SD), indicating growth delay. Owing to hypothyroidism, he has been prescribed 145 thyroxine. His motor development was delayed, with head control achieved at 6 months 146 of age, sitting at 14 months, and walking at 30 months. He forms no meaningful words, 147 and displays the most severe intellectual disability. Owing to poor eye contact, autistic 148 traits were also diagnosed. He can walk, but with an ataxic gait. He demonstrates 149 distinctive features, including a constant happy and smiling face, strabismus, a high forehead, a high and broad nasal bridge, and a protruding tongue. Owing to intractable 150

epilepsy, he has been prescribed anti-epileptic drugs. He requires support for his daily life. Comprehensive genomic analysis detected a nonsense variant in the lysine (K)-specific demethylase 5C gene (*KDM5C*) located on the X-chromosome, which was inherited from his carrier mother. Details of this variant have been reported previously ¹³⁾.

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III Discussion

The six chromosomal deletions including 13q31 band identified in this study were initially classified as "group 1" according to the classification scheme proposed by Brown et al. 1). Patient 3 showed congenital deafness and abnormal iris pigmentation. Because the deletion identified in patient 3 included the endothelin receptor type B gene (EDNRB), he was considered to have clinical features of Waardenburg syndrome type 4A (MIM # 277580) 14). The deletions identified in patients 1 and 2 also had microdeletions that included EDNRB. Patient 1 showed weak pigmentation of the iris, while patient 3 did not. These findings can be explained by variable expressions or incomplete penetrance of the clinical features ¹⁵⁾. Although patients with Waardenburg syndrome often also have Hirschsprung disease, patients 1-3 whose deletions included EDRNB displayed no clinical features of Hirschsprung disease. Puffenberger et al. suggested that the chance of developing Hirschsprung disease was 74% in homozygotes and 21% in heterozygotes ¹⁶⁾. Because *EDRNB* does not correlate with developmental delay, we considered that other candidate gene(s) would exist in this region. Regarding genotype-phenotype correlation in other patients, only the deletion identified in patient 5 extended to the 13q32.3 region, involving ZIC2 within the deletion region. This is why patient 5 showed holoprosencephaly and severe

developmental delay. Thus, this deletion may be classified as "group 2" rather than "group 1." Patient 6 had a small deletion in 13q31, which was inherited from his healthy father, indicating that the deletion identified in patient 6 was not responsible for his clinical features. Rather, patient 6 had a KDM5C mutation inherited from his mother, which could explain his clinical features. This finding gave us the important insight that haploinsufficiency of the genes included in the deletion identified in patient 6 would not underly clinical features such as neurodevelopmental delay. Jia et al. reported a similar small deletion in a healthy mother and child ¹⁷⁾. Thus, this region should also be excluded from the possible critical region for neurodevelopmental delay (Figure 1). Valdes-Miranda et al. reported a microdeletion involving 13q31.3-q32.1, identified in an individual with normal intelligence ¹⁸⁾. This entire deletion was included in the deletion in patient 4, narrowing the critical region for developmental delay to the distal region of patient 4's deletion. This region includes six genes (PGC5, GPC6, DCT, TGDC, GPR180, and SOX21), but none of them display a clear association with the neurodevelopmental delay observed in patient 4. Previously, Poreau et al. reported a fetus with a 13q31 deletion. Because the fetus was aborted, it is unknown whether this deletion was associated to neurodevelopmental delay ¹⁹⁾. As an alternative strategy, they analyzed the data included in DECIPHER database (https://www.deciphergenomics.org/). DECIPHER #256224 (chr13:79728762-79945691) showed language delay, but the identified microdeletion was inherited from the healthy parent. This small deletion encompasses only the RNA binding motif protein 26 gene (RBM26), suggesting that haploinsufficiency of RBM26 is likely benign and not responsible for neurodevelopmental delay. DECIPHER #268351 (chr13:79568105-80247708) showed language delay, and the identified microdeletion was inherited from

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a parent with a similar phenotype. Thus, a contribution of this deletion region to the clinical phenotype cannot be ruled out.

The result of the above discussion for genomic regions for neurodevelopmental delay narrowed the shortest region of overlapped (SRO) among patients 1-3 into (chr13:75691448_83625667) (Figure 1). Table 2 lists the genes included in the SRO and describes their relationships to their functions and phenotypes. Among them, the F-box and leucine-rich repeat protein 3 gene (*FBXL3*) is the only gene related to intellectual development, and mutations are associated with short stature, facial anomalies, and speech defects (MIM #606220). However, this condition displays autosomal recessive inheritance ²⁰⁾. Thus, haploinsufficiency of *FBXL3* would not be related to neurodevelopmental delay observed in patients with microdeletions in this region.

In the genes included in the SRO, only *MYCBP2* and *RBM26* show the probability of loss-of-function intolerance (pLI) score of "1" according to the gnomAD browser (https://gnomad.broadinstitute.org/). This indicates that loss-of-function of these genes may not be tolerated. The function of the RNA binding motif protein 26 (*RBM26*) is unknown but the MYC binding protein 2 E3 ubiquitin protein ligase gene (*MYCBP2*) may be related to neurodevelopmental delay, because *MYCBP2* regulates neuronal functions ²¹). Thus, *MYCBP2* haploinsufficiency may correlate with neurodevelopmental delay observed in patients 1-3.

In conclusion, we identified six microdeletions that include 13q31. Some regions were excluded from possible candidacy for neurodevelopmental delay. Because three patients (patients 1-3) with microdeletions including the proximal 13q31 region showed mild to moderate delay, some of the genes included in this deletion region are likely related to neurodevelopment. Distinctive features commonly observed in reported

223 patients were not detected. 224 We appreciate the cooperation of the patients and their families for this study. This 225 work was supported by JSPS KAKENHI JP18K07803, a grant from Health, Labour and 226 Welfare Sciences Research from MHLW Japan, and Initiative on Rare and Undiagnosed 227 Diseases (grant number 20ek0109301) from the Japan Agency for Medical Research 228 and Development (AMED). 229 230 **Conflict of interest** 231 All authors have no conflict of interest. 232

References

- 234 1) Brown S, Gersen S, Anyane-Yeboa K, Warburton D. Preliminary definition of a
- "critical region" of chromosome 13 in q32: report of 14 cases with 13q deletions
- and review of the literature. *Am J Med Genet* 1993;**45**:52-9.
- 237 2) Brown S, Russo J, Chitayat D, Warburton D. The 13q-syndrome: the molecular
- definition of a critical deletion region in band 13q32. Am J Hum Genet
- 239 1995;**5**7:859-66.
- 240 3) Brown SA, Warburton D, Brown LY, et al. Holoprosencephaly due to mutations in
- ZIC2, a homologue of Drosophila odd-paired. *Nat Genet* 1998;**20**:180-3.
- 4) Mimaki M, Shiihara T, Watanabe M, et al. Holoprosencephaly with cerebellar
- vermis hypoplasia in 13q deletion syndrome: Critical region for cerebellar
- 244 dysgenesis within 13q32.2q34. *Brain Dev* 2015;**37**:714-8.
- 245 5) Baud O, Cormier-Daire V, Lyonnet S, Desjardins L, Turleau C, Doz F. Dysmorphic
- phenotype and neurological impairment in 22 retinoblastoma patients with
- constitutional cytogenetic 13q deletion. *Clin Genet* 1999;**55**:478-82.
- 248 6) Skrypnyk C, Bartsch O. Retinoblastoma, pinealoma, and mild overgrowth in a boy
- with a deletion of RB1 and neighbor genes on chromosome 13q14. Am J Med
- 250 Genet A 2004;**124a**:397-401.
- 251 7) Ballarati L, Rossi E, Bonati MT, et al. 13q Deletion and central nervous system
- anomalies: further insights from karyotype-phenotype analyses of 14 patients. J
- 253 *Med Genet* 2007;44:e60.
- 8) Bellucco FT, Rodrigues de Oliveira-Júnior H, Santos Guilherme R, et al. Deletion
- of Chromosome 13 due to Different Rearrangements and Impact on Phenotype. *Mol*
- 256 *Syndromol* 2019;**10**:139-46.

- 257 9) Kirchhoff M, Bisgaard AM, Stoeva R, et al. Phenotype and 244k array-CGH
- characterization of chromosome 13q deletions: an update of the phenotypic map of
- 259 13q21.1-qter. Am J Med Genet A 2009;**149a**:894-905.
- 260 10) Quélin C, Bendavid C, Dubourg C, et al. Twelve new patients with 13q deletion
- syndrome: genotype-phenotype analyses in progress. Eur J Med Genet 2009;52:41-
- 262 6.
- 263 11) Shimojima K, Yamamoto T. Characteristics of rare and private deletions identified
- in phenotypically normal individuals. *Hum Genome Var* 2017;**4**:17037.
- 265 12) Fallet-Bianco C. Neuropathology of holoprosencephaly. Am J Med Genet C Semin
- 266 *Med Genet* 2018;**178**:214-28.
- 267 13) Yamamoto T, Imaizumi T, Yamamoto-Shimojima K, et al. Genomic backgrounds of
- Japanese patients with undiagnosed neurodevelopmental disorders. Brain Dev
- 269 2019;41:776-82.
- 270 14) Tüysüz B, Collin A, Arapoğlu M, Suyugül N. Clinical variability of Waardenburg-
- Shah syndrome in patients with proximal 13q deletion syndrome including the
- endothelin-B receptor locus. Am J Med Genet A 2009;149a:2290-5.
- 273 15) Pingault V, Bondurand N, Lemort N, et al. A heterozygous endothelin 3 mutation in
- Waardenburg-Hirschsprung disease: is there a dosage effect of EDN3/EDNRB gene
- mutations on neurocristopathy phenotypes? *J Med Genet* 2001;**38**:205-9.
- 276 16) Puffenberger EG, Kauffman ER, Bolk S, et al. Identity-by-descent and association
- 277 mapping of a recessive gene for Hirschsprung disease on human chromosome
- 278 13q22. Hum Mol Genet 1994;3:1217-25.
- 279 17) Jia Y, Zhao H, Shi D, et al. Genetic effects of a 13g31.1 microdeletion detected by
- 280 noninvasive prenatal testing (NIPT). *Int J Clin Exp Pathol* 2014;7:7003-11.

281	18) Valdes-Miranda JM, Soto-Alvarez JR, Toral-Lopez J, et al. A novel microdeletion
282	involving the 13q31.3-q32.1 region in a patient with normal intelligence. Eur J Med
283	Genet 2014; 57 :60-4.
284	19) Poreau B, Lin S, Bosson C, et al. 13q31.1 microdeletion: A prenatal case report
285	with macrocephaly and macroglossia. Eur J Med Genet 2015;58:526-30.
286	20) Ansar M, Paracha SA, Serretti A, et al. Biallelic variants in FBXL3 cause
287	intellectual disability, delayed motor development and short stature. Hum Mol
288	Genet 2019; 28 :972-9.
289	21) Holland S, Scholich K. Regulation of neuronal functions by the E3-ubiquitinligase
290	protein associated with MYC (MYCBP2). Commun Integr Biol 2011;4:513-5.
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294	Figure legends
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296	Table 1: Clinical features of the patients in this study
297	m, months; y, years; NT, not tested; w, weeks; d, days; NA, not available
298	
299	Table 2: List of the genes located on the possible candidate region
300	pLI, probability of loss-of-function intolerance; MIM, mendelian inheritance in man;
301	AR, autosomal recessive; AD, autosomal dominant
302	
303	Figure 1: Genome map of the region surrounding 13q31
304	Identified microdeletions are depicted on the genome map created by the UCSC genome
305	browser (https://www.genome.ucsc.edu/) using a custom track in accordance with
306	GRCh37/hg19. Grey and black bars with white arrow heads inside indicate deletions
307	identified in this study and the previously reported deletion (DECIPHER data are also
308	included), respectively. A translucent box shown by a black dotted line indicates the
309	shortest region overlapped discussed in the text.
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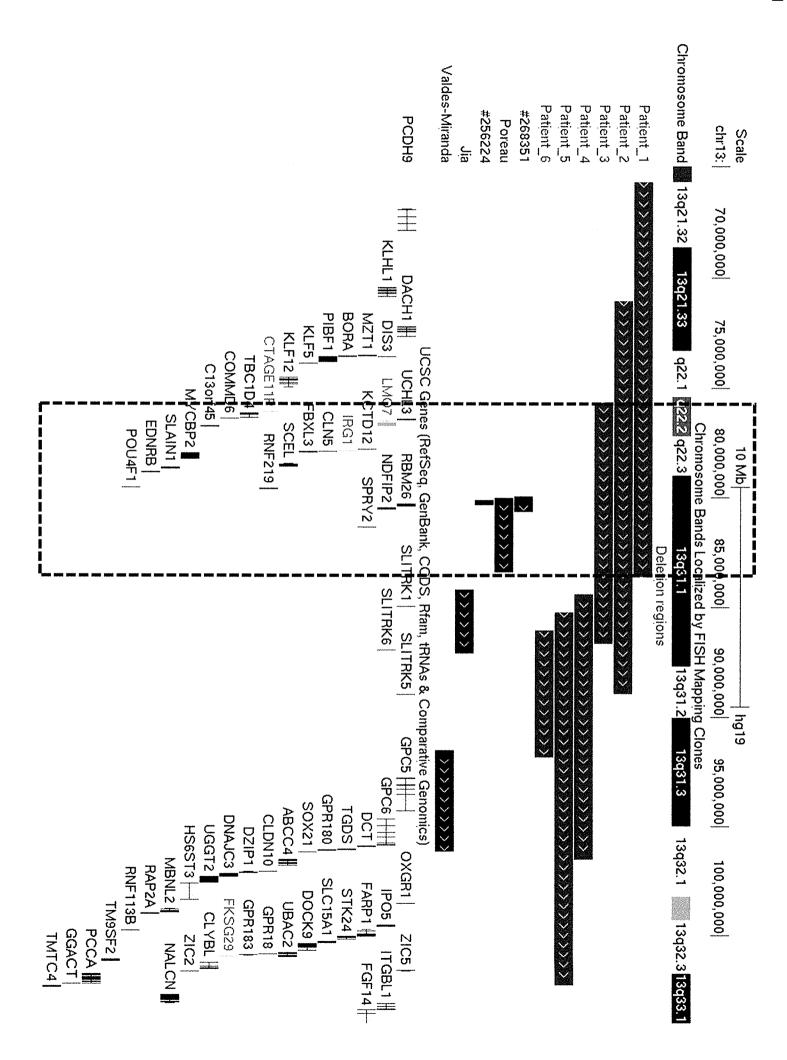


Table 1. Clinical features of the patients in this study

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- - - - aly Hypothyroidism		puberty,			paralysis	
1 1 1 1	Holoprosencepha	Precocious			Vocal cord	Other findings
1 1	1	ı	+	ı	ı	Wide ala nasi
1 1		ı	+	i	ı	Large auricle
1	+	1	ı	1	,	Flat nasal bridge
	•	ı	1	+	f	Micrognathia
ı	+	1	ı	ſ		Blepharophimosis
1		ı	+	ı	1	Epicanthus
ı	+	1	1	+	ı	Low-set-ears
						Distinctive facia features
Hypotonia	•		Hypotonia	1	ı	Neurological findings
ı	•	1	+	ı	ı	Deafness
2	r	ı		1	1	Hirschsprung disease
ı	•	ŧ	+	ŀ	+	Pigmentary changes of the
1	+	ı	+	ì	N A	Microcephaly
+	+	+	Mild	+	+	Growth deficiency
Moderate	Severe	Mild	Border	Moderate	+	Developmental delay
33.0	33.5	NA	33.0	29.8	30.3	Birth OFC
49.0	44.5	NA	49.2	43.9	47.4	Birth length
2556	2114	2284	3210	2555	2676	Birth weight
38w	39w2d	39w	38w1d	40w6d	39w6d	Gestational age
Inherited from father	N T	NT	de novo	Z	Z	Origin
5.8	17.0	12.1	11.0	17.9	17.9	Deletion Size [Mb]
91816350	102244722	96530663	86658563	88958577	83625667	End(bp)
86046386	85236169	84431421	75691448	71028436	65732480	Start(bp)
q31.1q31.3	q31.1q33.1	q31.1q32.1	q22.2q31.1	q21.33q31.2	q21.32q31.1	Chromosome band
60K	60K	60K	60K	60K	44K	Microarray kit
Male	Female	Female	Male	Male	Female	Sex
9у	18m	11y11m	8m	4y5m	3m	Age
Patient 6	Patient 5	Patient 4	Patient 3	Patient 2	Patient 1	

m, months; y, years; NT, not tested; w, weeks; d, days; NA, not available

Table 2. List of the genes located on the shortest region of overlapping (chr13:75691448_83625667)

	i ogion or or a	indepuis Com rous	10.10001	000	10001			
Gene name	Symbol	Start	End	рLI	Gene/Locus MIM number	Phenotype	Phenotype MIM number	Inheritance
CTAGE family, member 11, pseudogene TBC1 domain family, member 4	CTAGE11P TBC1D4	75811889 75858809	75814517 76056250	0	612465	Diabetes mellitus, noninsulin-dependent, 5	616087	
COMM domain containing 6	COMMD6	76099350	76111991	0.17	612377			
carboxyl-terminal esterase L3 (ubiquitin	UCHL3	76123886	76180156	0	603090			
thiolesterase)	I MOZ	76210459	76434006	>	604362			
chromosome 13 open reading frame 45	C13orf45	76445174	76457948	0 (0			
potassium channel tetramerisation domain	KCTD12	77454304	77460540	0.06	610521			
immunoresponsive 1 homolog (mouse)	IRG1	77526624	77532776	0	615275			
ceroid-lipofuscinosis, neuronal 5	CLN5	77566059	77576652	0	608102	Ceroid lipofuscinosis,	256731	AR
F-hox and leucine-rich reneat protein 3	ERYI 3	77579389	77601331	20 0	80555 8055 8055	neuronal, 5	606330	>
						developmental disorder with short stature, facial anomalies, and speech defects		
MYC binding protein 2, E3 ubiquitin protein	MYCBP2	77618792	77901177	_	610392			
ligase sciellin	SCEL	78109809	78219398	0	604112			
SLAIN motif family, member 1 endothelin receptor type B	SLAIN1 EDNRB	78271989 78469616	78338377 78493903	0.9	610491	Hirschsprung disease, susceptibility to, 2	600155	AD
						ABCD syndrome Waardenburg syndrome, type 4A	600501 277580	AR AD, AR
POU class 4 homeobox 1	POU4F1	79173230	79177695	0.89	601632			
RNA binding motif protein 26	RBM26	79894100	79979923	، د				
Nedd4 family interacting protein 2	NDFIP2	80055259	80130212	0.53	610041			
sprouty homolog 2	SPRY2	80910112	80915086	0.97	602466	lgA nephropathy, susceptibility to 3	616818	AD

pLl, probability of loss-of-function intolerance; MIM, mendelian inheritance in man; AR, autosomal recessive; AD, autosomal dominant